



An estimation of Central Iberian Peninsula atmospheric $\delta^{13}\text{C}$ and water δD in the Upper Cretaceous using pyrolysis compound specific isotopic analysis (Py-CSIA) of a fossil conifer.

José A González-Pérez (1), Nicasio T Jiménez-Morillo (1), José M De la Rosa (1), Gonzalo Almendros (2), and Francisco J González-Vila (1)

(1) IRNAS-CSIC, Seville, Spain, (2) MNCN-CSIC, Madrid, Spain.

Frenelopsis is a frequently found genus of the Cretaceous floras adapted to dry, saline and in general to environmental conditions marked by severe water stress [1]. Stable isotope analysis of fossil organic materials can be used to infer palaeoenvironmental variables helpful to reconstruct plant paleohabitats [2]. In this study stable isotope analysis of organic fossil remains (FR) and humic fractions (FA, HA and humin) of *Frenelopsis oligiostomata* are studied in bulk (C, H, O, N IRMS) and in specific compounds released by pyrolysis (C, H, Py-CSIA).

Well preserved *F. oligiostomata* fossils were handpicked from a limestone included in compacted marls from Upper Cretaceous (Senonian c. 72 Mya) in Guadalix de la Sierra (Madrid, Spain) [3]. The fossils were decarbonated with 6M HCl. Humic substances were extracted from finely ground fossil remains (FR) by successive treatments with 0.1M Na₄P₂O₇ + NaOH [4]. The extract was acidified resulting into insoluble HA and soluble FA fractions. The HA and FA were purified as in [5] and [6] respectively. Bulk stable isotopic analysis ($\delta^{13}\text{C}$, δD , $\delta^{18}\text{O}$, $\delta^{15}\text{N}$ IRMS) was done in an elemental micro-analyser coupled to a continuous flow Delta V Advantage isotope ratio mass spectrometer (IRMS). Pyrolysis compound specific isotopic analysis Py-CSIA ($\delta^{13}\text{C}$, δD): was done by coupling a double-shot pyrolyzer to a chromatograph connected to an IRMS. Structural features of specific peaks were inferred by comparing/matching mass spectra from conventional Py-GC/MS (data not shown) with Py-GC/IRMS chromatograms obtained using the same chromatographic conditions.

Bulk C isotopic signature found for FR (-20.5 ± 0.02 ‰) was in accordance with previous studies [2, 7–9]. This heavy isotopic $\delta^{13}\text{C}$ signature indicates a depleted stomatal conductance and palaeoenvironmental growth conditions of water and salt stress. This is in line with the morphological and depositional characteristics [3] confirming that *F. oligiostomata* was adapted to highly xeric and saline habitats being a component of salt-marsh vegetation. The values obtained for δD (-101.9 ± 2.2 ‰), $\delta^{15}\text{N}$ (10.7 ± 0.2 ‰) and $\delta^{18}\text{O}$ (20.9 ± 0.39 ‰) lay within those previously reported for fossil floras [10] growing in warm environment and probably with very high evaporation rates.

$\delta^{13}\text{C}$ Py-CSIA was recorded for biogenic compound; polysaccharides, lipid series, lignin and degraded lignin compounds (alkyl benzenes and alkyl phenols) and for a S containing compounds probably with a diagenetic origin. In general $\delta^{13}\text{C}$ Py-CSIA values were more depleted than the bulk ones and can be considered a better approach to the real plant $\delta^{13}\text{C}$ value (c. -22 ‰). Considering that plant-air C fractionation in degraded lignin compounds for a C₄ photosystem plant is c. $\Delta^{13}\text{C} \approx 20.0$ ‰ [11] and an extra fractionation ($\Delta^{13}\text{C} \approx -3.0$ ‰) due to the plant depleted stomatal conductance growing in extreme warm, saline and dry conditions, we estimate atmospheric $\delta^{13}\text{C}$ value in the area during the Upper Cretaceous in c. $\delta^{13}\text{C} = -5.3 \pm 0.2$ ‰. This indicates that our *F. oligiostomata* probably grew on a ^{13}C enriched atmosphere, more enriched than preindustrial one ($\delta^{13}\text{C} \approx -6.5$ ‰ [12]). This could be caused by a combination of reasons, i.e. emission of heavy ^{13}C isotopes to the atmosphere by an increase in values for δD CSIA of lipid compounds such as n-alkanes with C chain lengths, C₂₃–C₃₁ are believed to derive exclusively from leaf waxes of higher plants. Plant δD carries isotope information of environmental water that is particularly preserved during the geological record in n-alkyl structures, whereas other structures i.e. isoprenoids, are most prone to hydrogen exchange [13–14]. We were able to measure δD for long chain alkane/alkene series in the range C₂₄–C₂₉ ($\delta\text{D} = -124.44 \pm 5.2$ ‰). This was taken as a proxy to infer the original H isotopic signal of water in the area in the Upper Cretaceous. Poole et al. (2004) proposed that $\delta\text{D}_{\text{palaeowater}} = \delta\text{D}_{\text{C}_{24}\text{--}\text{C}_{29}} - 100$ giving a value for palaeowater $\delta\text{D} = -24.44 \pm 5.2$ ‰. This indicates that 75 Mya our plant probably uptake deuterium enriched rain water that again points to warm growing environmental conditions.

(1) Gómez, B.; Martín-Closas C.; Brale G.; Solé de Porta N.; Thévenard F.; Guignard G. *Paleontology* 2002 45, 997–1036.

(2) Nguyen Tu, T.T.; Kvaček, J.; Uličný, D.; Bocherens, H.; Mariotti, A.; Broutin, J. *Palaeogeogr. Palaeoclimatol.*

Palaeoecol. 2002 183, 43–70.

(3) Almendros, G.; Álvarez-Ramis, C.; Polo, A. *Revista de la Real Academia de Ciencias Exactas, Físicas y Naturales* 1982 76, 285–302.

(4) Dabin, B. *Chah. ORSTOM Ser. Pedol.* 1976 4, 287–297.

(5) Schnitzer, M.; Khan, S.U. *Humic Substances in the Environment*. Marcel Dekker Inc. 1972, New York, N.Y.

(6) Dorado, E.; Polo, A. *An. Edafol. Agrobiol.* 1976 55, 723–732.

(7) Bocherens, H.; Friis, E.M.; Mariotti, A.; Pedersen, K.R. *Lethaia* 1993 26, 347–358.

(8) Nguyen Tu, T.T.; Bocherens, H.; Mariotti, A.; Baudin, F.; Pons, D.; Broutin, J.; Derenne, S.; Largeau C. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 1999 145, 79–93.

(9) Aucour, A.-M.; Gomez, B.; Sheppard, S.M.F., Thévenard, F. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 2008 257, 462–473.

(10) Michener, N.; Lajtha K. (Eds). *Stable Isotopes in Ecology and Environmental Science* (2nd Ed) 2007 Blackwell Publishing.

(11) Poole, I., van Bergen, P.F.; Kool, K.; Schouten, S.; Cantrill, D. *J. Org. Geochem.* 2004 35, 1261–1274.

(12) Gerber, S.; Joos, F.; Brügger, P.; Stocker, T.F.; Mann, M.E.; Sitch, S.; Scholze, M. *Clim. Dyn.* 2003 20, 281–299, 2003

(13) Pedentchouk, N.; Freeman, K.H.; Harris, N.B. *Geochim. Cosmochim. Acta* 2006 70, 2063–2072.

(14) Radke, J.; Bechtel, A.; Gaupp, R.; Püttmann, W.; Schwark, L.; Sachse D.; Gleixner, G. *Geochim. Cosmochim. Acta* 2005 69, 5517–5530.

Acknowledgements

Projects CGL2012-38655-C04-01 and CGL2008-04296 and fellowship BES-2013-062573 given by the Spanish Ministry for Economy and Competitiveness to N.T.J.M. Dr. J.M. R. is the recipient of a fellowship from the JAE-Doc subprogram financed by the CSIC and the European Social Fund.