

## Estimating N<sub>2</sub>O processes during grassland renewal and grassland conversion to maize cropping using N<sub>2</sub>O isotopocules

Caroline Buchen, Reinhard Well, Heinz Flessa, Roland Fuß, Mirjam Helfrich, and Dominika Lewicka-Szczebak  
Thünen Institute, Climate-Smart Agriculture, Braunschweig, Germany (reinhard.well@thuenen.de)

Grassland break-up due to grassland renewal and grassland conversion to cropland can lead to a flush of mineral nitrogen from decomposition of the old grass sward and the decomposition of soil organic matter. Moreover, increased carbon and nitrogen mineralisation can result in enhanced nitrous oxide (N<sub>2</sub>O) emissions. As N<sub>2</sub>O is known to be an important greenhouse gas and a major precursor for ozone depletion, its emissions need to be mitigated by adjusting agricultural management practices. Therefore, it is necessary to understand the N<sub>2</sub>O processes involved, as well as the contribution of N<sub>2</sub>O reduction to N<sub>2</sub>. Apart from the widely used <sup>15</sup>N gas flux method, natural abundance isotopic analysis of the four most abundant isotopocules of N<sub>2</sub>O species is a promising alternative to assess N<sub>2</sub>O production pathways.

We used stable isotope analyses of soil-emitted N<sub>2</sub>O ( $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ ,  $\delta^{15}\text{N}_{\text{N}_2\text{O}}^{\text{bulk}}$  and  $\delta^{15}\text{N}_{\text{N}_2\text{O}}^{\text{SP}}$  = intramolecular distribution of <sup>15</sup>N within the linear N<sub>2</sub>O molecule) with an isotopocule mapping approach to simultaneously estimate the magnitude of N<sub>2</sub>O reduction to N<sub>2</sub> and the fraction of N<sub>2</sub>O originating from the bacterial denitrification pathway or fungal denitrification and/or nitrification. This approach is based on endmember areas of isotopic values for the N<sub>2</sub>O produced from different sources reported in the literature. For this purpose, we calculated two main scenarios with different assumptions for N<sub>2</sub>O produced: N<sub>2</sub>O is reduced to N<sub>2</sub> before residual N<sub>2</sub>O is mixed with N<sub>2</sub>O of various sources (Scenario a) and vice versa (Scenario b). Based on this, we applied seven different scenario variations, where we evaluated the range of possible values for the potential N<sub>2</sub>O production pathways (heterotrophic bacterial denitrification and/or nitrifier denitrification and fungal denitrification and/or nitrification). This was done by using a range of isotopic endmember values and assuming different fractionation factors of N<sub>2</sub>O reduction in order to find the most reliable scenario. Investigations were carried out over a study period of one year following grassland renewal and grassland conversion to maize cropping on two different soil sites (Plaggic Anthrosol and Histic Gleysol) near Oldenburg, Lower Saxony Germany.

Our observations indicate heterotrophic bacterial denitrification and/or nitrifier denitrification as the main source of N<sub>2</sub>O production, with a significant contribution of N<sub>2</sub>O reduction to N<sub>2</sub> rather than nitrification (i.e. hydroxylamine oxidation) and fungal denitrification throughout the entire study period. A tendency to a higher contribution of N<sub>2</sub>O reduction to N<sub>2</sub> was observed for the often water-saturated Histic Gleysol, while lower N<sub>2</sub>O reduction was found for the Plaggic Anthrosol. For two samples, we attempt to validate our results from the isotopocule mapping approach with a parallel <sup>15</sup>N labelling study at the field scale (Buchen et al., 2016), as conditions of soil moisture, nitrate availability and N<sub>2</sub>O flux were similar.

### References:

Buchen, C., Lewicka-Szczebak, D., Fuß, R., Helfrich, M., Flessa, H., Well, R., 2016. Fluxes of N<sub>2</sub> and N<sub>2</sub>O and contributing processes in summer after grassland renewal and grassland conversion to maize cropping on a Plaggic Anthrosol and a Histic Gleysol. *Soil Biology and Biochemistry* 101, 6-19.