



The response of soil N cycling and nitrous oxide emission to Free Air CO₂ Enrichment in a temperate forest, UK.

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Increasing atmospheric CO₂ concentrations in temperate forests may affect soil nitrogen (N) cycling processes due to the increased demand for nitrogen availability to support CO₂ uptake by trees through photosynthesis. This in turn can affect the emission of nitrous oxide (N₂O), a potent greenhouse gas, from the forest soil leading to a potential trade-off between the enhanced canopy CO₂ uptake and soil N₂O emission. Our current understanding of the response of N cycling processes and availability to elevated atmospheric CO₂ (eCO₂) in mature, unmanaged temperate forests is limited. The Birmingham Institute of Forest Research (BIFoR) established in 2017 a Free-Air CO₂ Enrichment (FACE) facility in a mature temperate oak dominated forest in Staffordshire, UK, to study under 'real world' conditions the risks of a changing climate to forest ecosystems and the services they provide. In April 2018, one year after the start of fumigation with 550 ppm CO₂, we collected soil samples (0 – 15 cm depth) from the three eCO₂ and three control plots. Being a N limited forest soil in spite of enhanced atmospheric reactive N deposition (22 kg N ha⁻¹ y⁻¹), we hypothesized that N mineralisation will increase and N₂O emission will be down regulated to meet tree N demands under eCO₂. Soils were amended in the laboratory with 98 at % ¹⁵N-NH₄⁺ and ¹⁵N-NO₃⁻ (at ~ 20 % of the ambient soil NH₄⁺ and NO₃⁻ concentration) and were incubated in the dark for 24 hours. Gross N mineralisation and nitrification were estimated according to the isotope dilution technique, while N₂O emission from nitrification (¹⁵N-NH₄⁺ treatment) and denitrification (¹⁵N-NO₃⁻ treatment) were estimated according to the ¹⁵N Gas-Flux method. Additionally, asymbiotic biological N fixation was measured with the ¹⁵N₂ direct assimilation method. Gross N mineralisation was significantly higher (t-Test: $P < 0.05$) in the eCO₂ plots (mean: 2.25 μg N g⁻¹ d⁻¹) compared to the control plots (mean: 0.61 μg N g⁻¹ d⁻¹) confirming our hypothesis, while there was no significant difference in gross nitrification rates. N₂O emission from both denitrification (mean: 2.85 ng N g⁻¹ d⁻¹) and nitrification (mean: 0.18 ng N g⁻¹ d⁻¹) was marginally higher in the eCO₂ plots, but the difference was not statistically significant (t-Test: $P > 0.05$). Denitrification was a stronger source of N₂O (16 times higher than nitrification) in the eCO₂ plots compared to the controls (denitrification 12 times higher than nitrification). Finally, asymbiotic biological nitrogen fixation was also significantly higher in eCO₂ plots. After one year of CO₂ fumigation, there is indication of enhanced cycling and N availability to support enhanced canopy CO₂ uptake. As more soil inorganic N becomes available a shift towards higher N₂O emission is to be expected and continuous multiannual *in situ* monitoring is needed to establish the balance between N availability for tree uptake relative to dissimilatory N losses including N₂O emission under eCO₂ to fully elucidate the implications of N availability for meeting tree N demands under future CO₂ enriched atmosphere.