Nicrobial mechanisms

RE

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The concept of biomineralization distinguishes **microbially controlled** and **microbially induced** formation; both may be intracellular or extracellular

Our research on heavy metal resistance mechanisms in bacteria and fungi led us to consider a more microbiological distinction based rather on **metabolic active** or **passive** processes



Crystal formation is dependent on living biomass

Streptmyces acidiscabies forms nickel struvite crystals surrounded by the bacterial mycelium. Autoclaved biomass does not lead to crystal formation





Different strains form different crystals



Habit of single crystals of Ni(NH₄)(PO₄)·6H₂O A) possible habit formed by {00-1}, {100}, {010}, {113} and {001} faces B) view direction parallel to [010] of an idealized struvite crystal formed by {010}, {100}, {103} and {10-3} faces



Active pH control leads to different minerals formed

Streptmyces lividans TK 23 acidifies the medium

Streptmyces naganishii P9A-1 leads to alkalinization (without urea in the medium!)





Minerals at different distance to the colony

Calcit (Calcium carb<u>onate</u>, Sphingopyxis bauzanensis

Vaterit (Calcium carbonate, *Moraxella osloensis* sMM16)



Genetic determinants



A megaplasmid of *Streptomyces mirabilis* P16-B1 with several metal resistance genes leads to enhanced biomineral formation if transferred to a metal sensitive S. lividans.

The plasmid is naturally transferable in soil microcosms



1.0 mm

Bioweathering and biomineralization are connected

The basidiomycete fungus *Schizophyllum commune* can degrade shale (by laccase involvement)

This releases metals which induce metal resistance gene (as shown with transcriptome and proteome studies



Oxalates are formed, likely connected to extracellular matrix (the glucan schizophyllan could be shown)





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"Alteration and element mobility at the microbe-mineral interface"

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